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URINARY METHYLMALONIC ACID AS AN INDEX OF
VITAMIN B₁₂ METABOLISM

by

JOAN ELIZABETH SUTHERLAND DAKIN

A THESIS

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The undersigned certify that they have read, and recommend to the Faculty of Graduate Studies for acceptance, a thesis entitled "METHYLMALONIC ACID EXCRETION AS AN INDEX OF VITAMIN B₁₂ DEFICIENCY", submitted by Joan Elizabeth Sutherland Dakin in partial fulfillment of the requirements for the degree of Master of Science.

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ABSTRACT

Methylmalonic acid, a metabolite of propionic acid, has been reported (1,2,3) to be increased in the urine of patients with untreated pernicious anemia. In order to study methylmalonic acid excretion and vitamin B₁₂ deficiency, a quantitative method for the determination of methylmalonic acid in urine was developed. The method uses the technique of thin-layer chromatography on silica gel of an ether extract of urine. The normal urinary excretion of methylmalonic acid by this method was determined to be less than 10 mg. per twenty-four hours.

In twenty-three cases of clinically proven untreated pernicious anemia, urinary levels of methylmalonic acid were found to be increased. The level of methylmalonic acid returned to normal in response to vitamin B₁₂ therapy. These cases also exhibited low Co⁶⁰-vitamin B₁₂ absorption. There appeared to be no correlation between the level of methylmalonic acid and the degree of neurological complication. In some cases, the urinary methylmalonic acid was elevated before hematological abnormalities developed.

As pernicious anemia from a biochemical point of view can be considered to be a defect in nucleoprotein synthesis, it was of interest to determine urinary levels

of 4(5)-amino-5(4)-imidazolecarboxamide nucleotide, an important intermediate in the synthesis of purine nucleotides. This was done in the patients with pernicious anemia and levels were found to be only occasionally elevated. This is at variance with reports (5,6) of consistent elevations of urinary 4(5)-amino-5(4)-imidazolecarboxamide in vitamin B₁₂ deficiency.

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I. INTRODUCTION

Pernicious anemia is a disease of vitamin B₁₂ deficiency. The classical signs of vitamin B₁₂ deficiency include megaloblastosis (due to arrested nucleoprotein synthesis), anemia, neuromyelopathy, encephalopathy, glossitis and alimentary disturbances. The laboratory findings include specific hematologic abnormalities (hypersegmentation of the polymorphonuclear neutrophils, macrocytosis, anemia and megaloblastic erythropoiesis as shown in the bone marrow), histamine-fast achlorhydria, low serum vitamin B₁₂ levels and a subnormal absorption of radio-active vitamin B₁₂ (Schilling test). The condition responds to injection of vitamin B₁₂, the reticulocyte count being a measure of the hematopoietic response.

In 1962, Cox and White (1) first reported the increased level of methylmalonic acid (MMA), a metabolite of propionic acid, in the urine of patients with pernicious anemia in relapse and suggested that this measurement might serve as an index of vitamin B₁₂ deficiency. They found that normal controls and patients with normal serum B₁₂ levels had MMA excretions not exceeding 3.8 mg. in twenty-four hours (2). In addition to the markedly increased urinary excretion of MMA in patients with pernicious anemia, it was found that all patients with low serum vitamin B₁₂ levels had an increased excretion of MMA, although this was less noticeable in

patients without a disordered erythropoiesis. It was found that, although a constant relationship exists between subnormal serum B₁₂ levels and increased urinary excretion of MMA, there is no proportionate relationship. These authors used a gas chromatographic separation coupled with isotope dilution to estimate urinary MMA.

In 1963, Barness (3) reported a urinary excretion of 500 mg. per day of MMA in one case of pernicious anemia. Following intramuscular doses of vitamin B₁₂, MMA could not be detected in the urine. Barness used two-dimensional paper chromatography to separate MMA from other urinary organic acids. Elution of the acid, followed by titration, gave quantitative results. The excretion level reported of 500 mg. per day in pernicious anemia is five times higher than the highest level reported by Cox and White in five cases of untreated pernicious anemia. This may suggest a lack of specificity in the paper chromatographic procedure.

In view of the interest in the excretion of MMA in vitamin B₁₂ deficiency, it was proposed to attempt to develop a simple method for the measurement of MMA in urine. The usefulness of this measurement as an index of vitamin B₁₂ deficiency could be partially assessed by comparison with serum vitamin B₁₂ levels and results obtained from Schilling tests. Also, Cox and White suggested that there may be a relation between the neurological complications of vitamin B₁₂ deficiency and the MMA excretion (reflecting elevated serum MMA). It was thought that the possibility of such a

relationship should also be investigated.

As further evidence of the interest in vitamin B₁₂ metabolism, there have been several reports (4,5,6) of increased urinary excretion of 4(5)-amino-5(4)-imidazole-carboxamide (AIC) in patients with vitamin B₁₂ and/or folic acid deficiency. AIC in the ribotide form is a key intermediate in the biosynthesis of the purine moiety of nucleic acids. It was, therefore, decided to measure urinary levels of AIC.

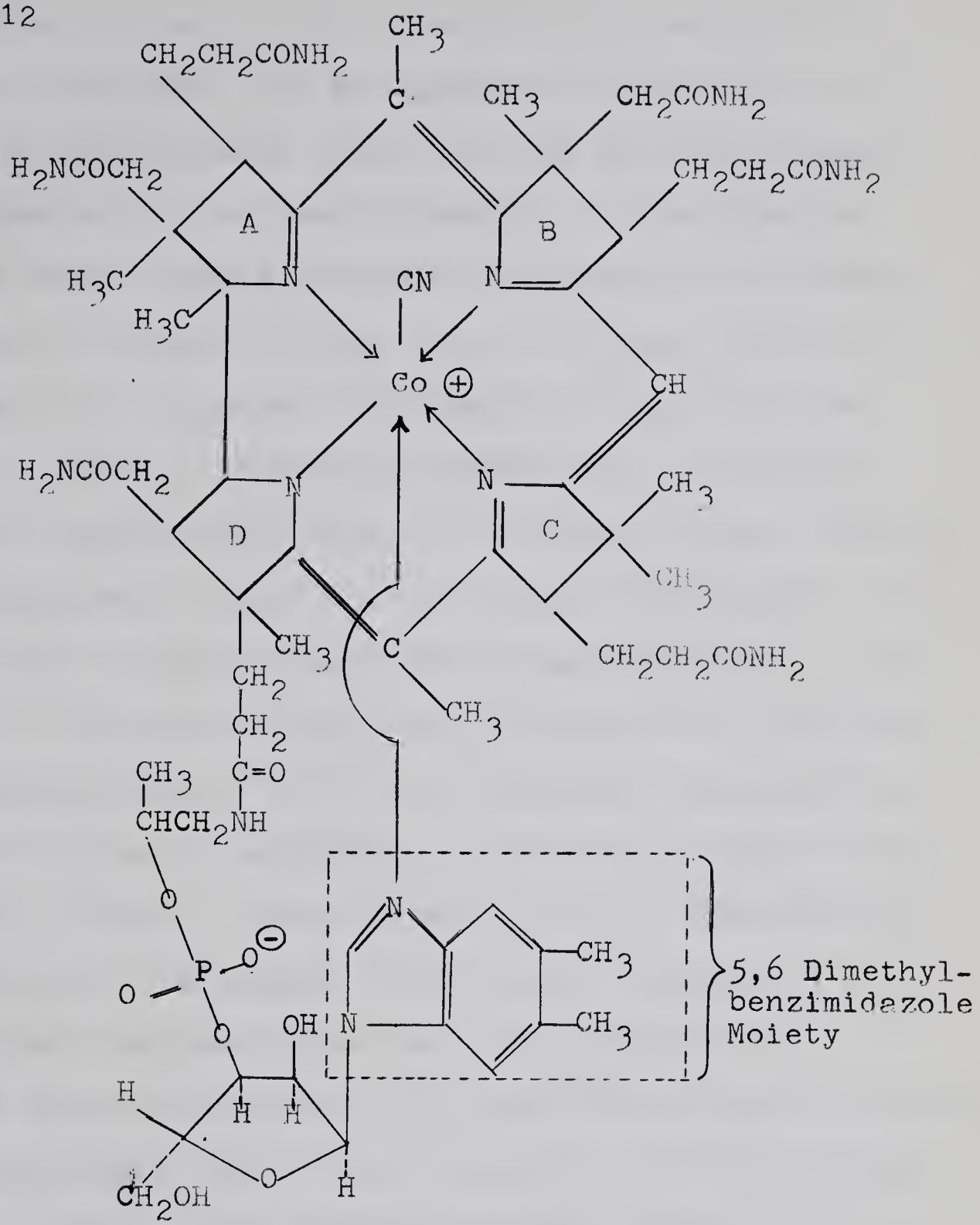
II. VITAMIN B₁₂

Vitamin B₁₂, the anti-pernicious anemia factor or extrinsic factor of Castle, was first isolated from liver in 1948. In 1958, Barker et al (7) isolated three vitamin B₁₂ containing coenzymes from microbial sources. Vitamin B₁₂ is unique among the vitamins in that it is synthesized only by microorganisms.

A. CHEMISTRY

Vitamin B₁₂, C₆₃H₈₈O₁₄N₁₄PCo, molecular weight 1356, is a red crystalline compound and has a structure as shown in Figure I. It consists of two major structures, a planar group resembling a porphyrin ring, and a nucleotide nearly perpendicular to the planar group. The planar group consists of four reduced and extensively substituted pyrrole rings (A - D) linked to a central cobalt atom. The pyrrole rings are connected to one another by methene bridges except for a direct linkage between the α carbons of rings A and D.

VITAMIN B₁₂



COENZYME

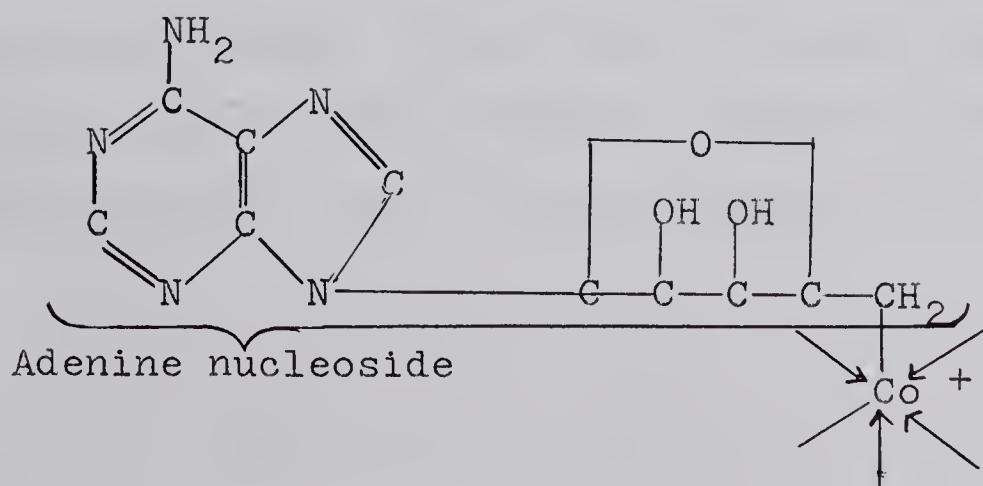


FIGURE I. Structure of Vitamin B₁₂ and the B₁₂ coenzyme.

Below the planar group is the nucleotide, 5,6,-dimethylbenzimidazole ribotide. It is connected at one end in a coordinate linkage between cobalt and one of the nitrogens of the imidazole ring of benzimidazole. At the other end there is an ester linkage between the nucleotide phosphate and a 1-amino-2-propanol moiety that is in turn joined in amide linkage to a propionic side chain on ring D of the tetrapyrrole nucleus. Above the planar group, a cyanide group is also coordinately bound to the cobalt atom. Removal of the cyanide group results in a compound "cobalamin". The presence of the cyanide group forms "cyanocobalamin". Substitution of the cyanide group with a hydroxy or nitro group forms "hydroxycobalamin" and "nitrocobalamin" respectively.

The cobamide coenzymes are the biologically active forms of the vitamin. The coenzymes have a 5'-deoxyadenosyl group in place of the cyanide ligand above the plane. The three cobamide coenzymes which have been identified all possess the adenine nucleoside but they differ from one another in the benzimidazole part of the molecule. In this position there may be the 5,6,-dimethylbenzimidazole group as in vitamin B₁₂, an unsubstituted (methyl free) benzimidazole group or an adenyl group. These three cobamide coenzymes are designated 5,6,-dimethylbenzimidazole cobamide (DBC), benzimidazole cobamide (BC) or adenyl cobamide (AC) coenzymes.

B. PHYSIOLOGY

In man, the whole body pool size of vitamin B₁₂ is approximately 5 mg. and the metabolic turnover rate about 2.5 μ g. per day (8). Clinical symptoms of vitamin B₁₂ deficiency do not appear until the body pool is depleted to about 10% of the normal value.

Vitamin B₁₂ is absorbed from the ileum and the absorption is dependent upon the presence of intrinsic factor (IF). IF is secreted by the fundus of the normal stomach and while it has not yet been isolated in pure form is thought to be a mucopolypeptide or mucopolysaccharide. The mechanisms of the vitamin B₁₂-IF binding and the absorption of vitamin B₁₂ are not known. It has been suggested (9) that there are "receptors" for either IF alone or the vitamin B₁₂-IF complex in the wall of the ileal mucosa, and that there may be two binding sites on the IF molecule, one for vitamin B₁₂ and one for the "receptor" in the gut wall. Large molecular complexes, when absorbed, are transported via the lymphatics. Since absorbed vitamin B₁₂ travels in the blood stream it is probably absorbed alone rather than in combination with IF. This would suggest a system in the ileum or ileal mucosa that releases vitamin B₁₂ from IF. Vitamin B₁₂ is transported in the blood stream, 80 - 85% being bound to an alpha globulin, and is stored in the liver.

Megaloblastic anemia is the hematologic result of any of a variety of etiologic factors causing improper

nucleoprotein synthesis. The most common causes are deficiencies of vitamin B₁₂, folate, or both. Vitamin B₁₂ deficiency has been generally classified by Herbert (10) as resulting from:

1. Inadequate ingestion, e.g. malnutrition, vegetarians, "vegans". In this case, 5 micrograms of vitamin B₁₂ orally per day is adequate therapy.

2. Inadequate absorption

a. Inadequate secretion of IF

In the absence of IF, practically none of the 2 - 10 micrograms of vitamin B₁₂ present in the average diet can be absorbed. The vitamin deficiency then develops over a three to six year period, the time taken to nearly exhaust the liver stores.

Addisonian pernicious anemia is due to inadequate IF secretion and is usually associated with gastric atrophy. Parenteral administration monthly of 250 or 500 micrograms of vitamin B₁₂ serves to treat the condition.

b. Functional abnormality of ileum

Vitamin B₁₂ deficiency may be due to inadequate absorption due to a functional or structural abnormality in the ileum, the main site of vitamin B₁₂ absorption.

c. Inadequate utilization

This cause of vitamin B₁₂ deficiency has not yet been demonstrated but, according to Herbert (10), will be the inevitable consequence of anti-vitamin B₁₂ therapy of cancer which has been proposed.

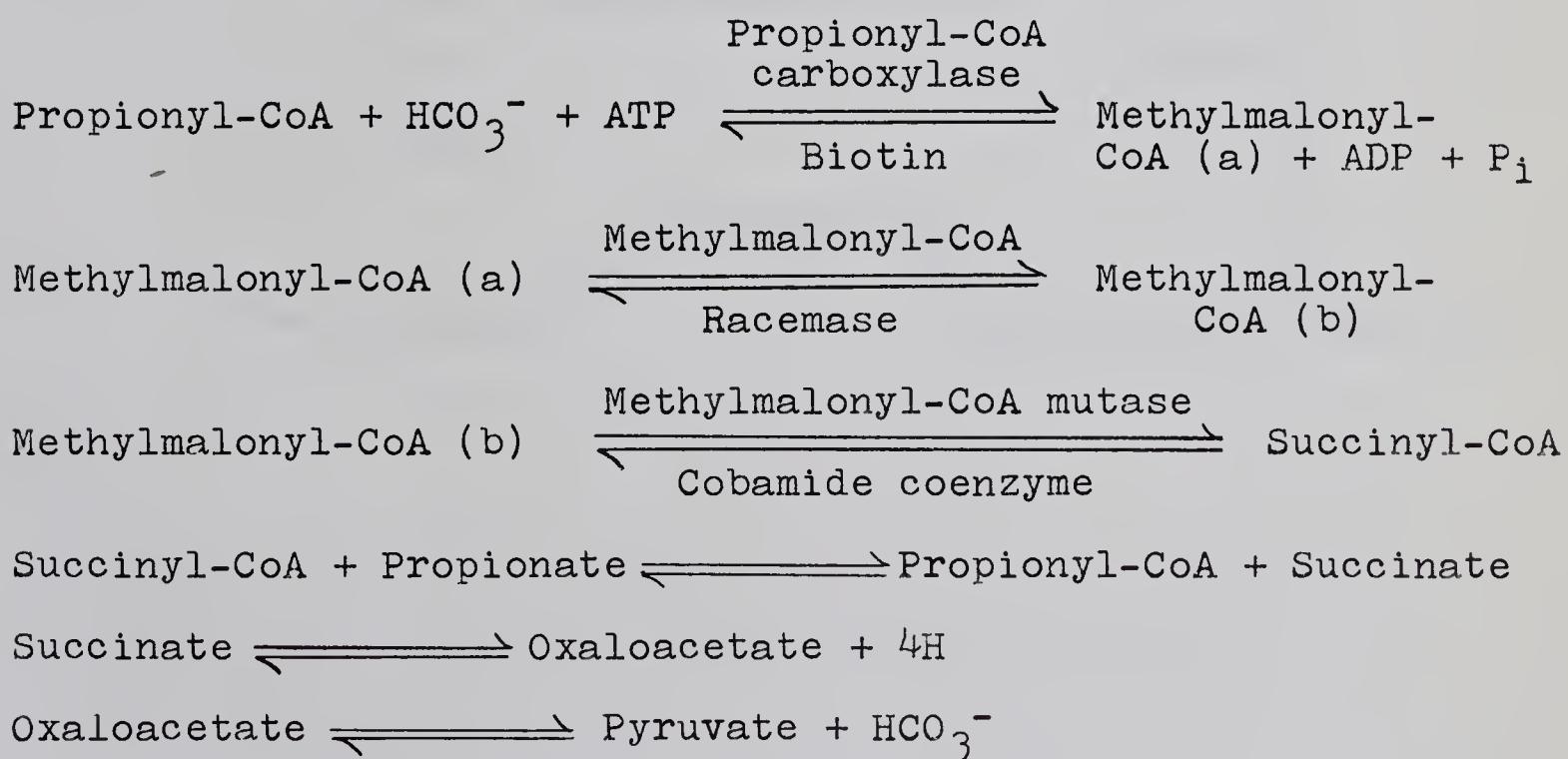
C. ROLE IN METABOLISM

Vitamin B₁₂ functions as a cobamide coenzyme in several reactions in animals and microorganisms. In addition, it is involved in DNA synthesis largely due to its interrelation with folic acid.

1. Enzymatic Reactions Involving Cobamide Coenzymes

a. Methylmalonyl - CoA mutase (isomerase)

In 1955, Flavin, Ortiz and Ochoa (11) showed that a major pathway of propionic acid utilization in tissues of animals involves the conversion of propionyl-CoA to succinyl-CoA by way of an intermediate compound methylmalonyl-CoA. The series of reactions which occur is shown in Figure II and can be summarized as follows:



SUM:



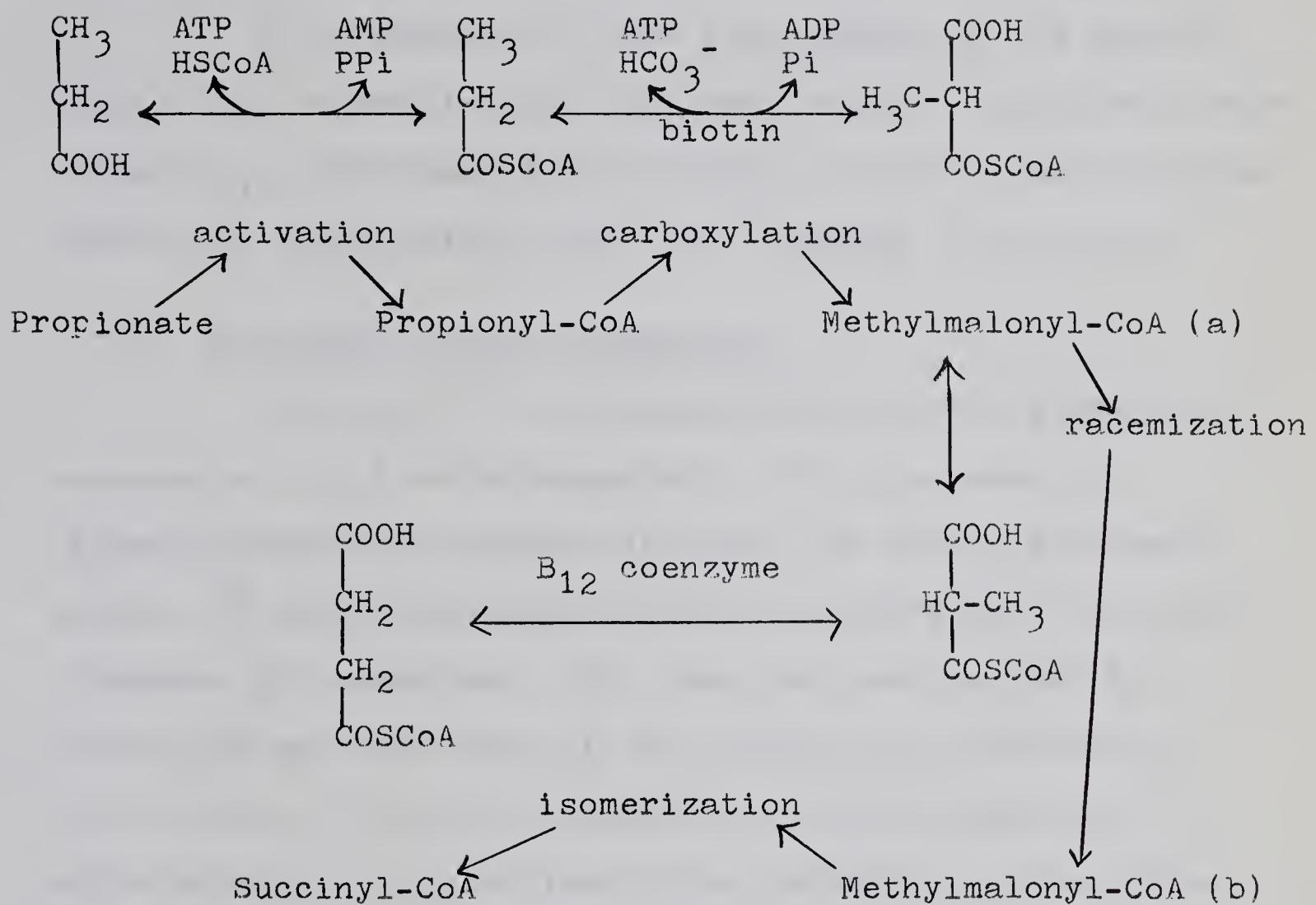


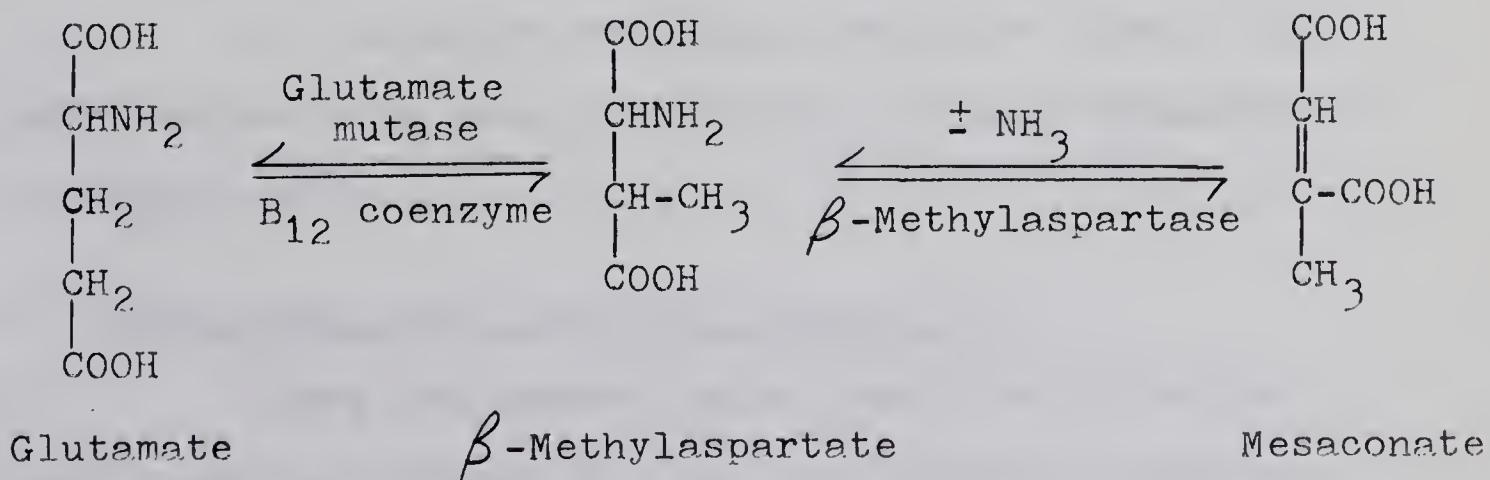
FIGURE II. Conversion of propionate to succinyl-CoA.

It was shown that dimethyl benzimidazole cobamide (DBC) coenzyme was involved in the isomerization of methylmalonyl-CoA (b) to succinyl-CoA. This isomerization occurs by an intramolecular shift of the CoA - carboxyl between the α and β carbons of the propionic acid moiety.

It is because of this requirement of the methylmalonyl-CoA mutase for the DBC coenzyme that in patients with vitamin B₁₂ deficiency this reaction is blocked and increased amounts of methylmalonic acid are excreted in the urine.

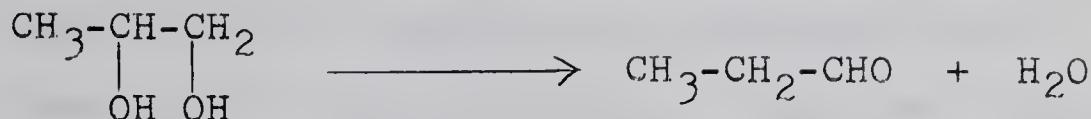
b. Glutamate mutase (isomerase)

Extracts of *Cl. tetanomorphum* convert glutamate to mesaconate via β -methylaspartate. The glutamate to β -methylaspartate reaction involves the enzyme glutamate mutase (β -methylaspartate mutase) which requires the adenylyl cobamide (AC) coenzyme. This coenzyme was isolated by Barker and was the first of the vitamin B₁₂ coenzymes to be isolated. It is the coenzyme of pseudovitamin B₁₂ in which adenine replaces dimethylbenzimidazole in the nucleotide moiety. The following are the reactions involved.



c. Dioldehydrase

Cell free extracts of *A. aerogenes* catalyze a cobamide dependent dehydration of glycols to the corresponding deoxyaldehyde, specifically 1,2 propanediol to propionaldehyde,



and ethylene glycol to acetaldehyde.



The enzyme, dioldehydrase, of *A. aerogenes* requires the AC coenzyme as cofactor.

d. Lysine degradation

C1. Sticklandii and *Clostridium ME* degrade lysine in the presence of pyruvate to yield one mole each of butyrate and acetate and two moles of ammonia. The reaction involves the participation of a cobamide coenzyme, is inhibited by intrinsic factor, leads to ATP formation and requires a flavin, a disulfide and Fe^{++} ion.

Of the above reactions, only the first, the methylmalonyl-CoA mutase reaction, has been demonstrated in animal cells.

2. Other Reactions Involving Vitamin B₁₂

There are several other reactions in which involvement of vitamin B₁₂ has been shown or postulated. Some of these reactions are ones which primarily involve folic acid.

a. Biosynthesis of methionine methyl group

Both vitamin B₁₂ and folic acid are implicated in the biosynthesis of the methyl group of methionine.

Homocysteine + N⁵-Methyl tetrahydrofolic acid



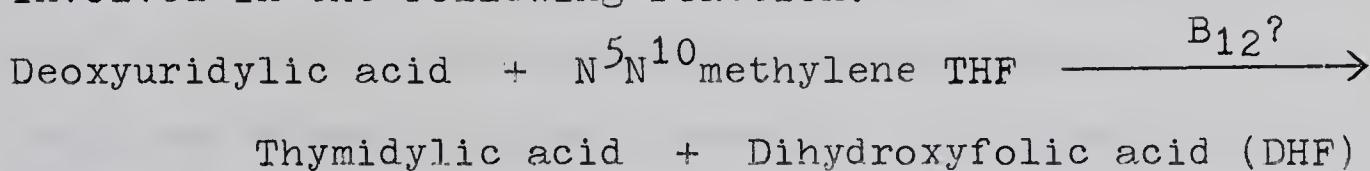
The enzyme 5-Methyl THF - homocysteine transmethylase catalyzes irreversibly the vitamin B₁₂ - dependent transfer of methyl groups with the formation of methionine and THF.

b. Deoxyribose synthesis

It has been suggested (13, 14) as a result of work with cultures of *L. leichmannii* that vitamin B₁₂ may participate in the conversion of ribose to deoxyribose.

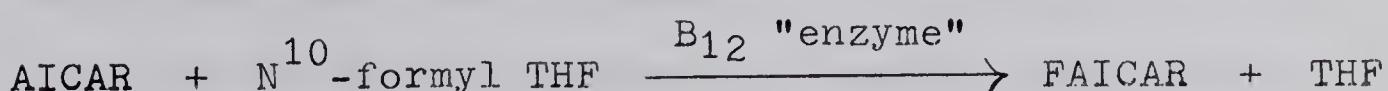
c. Thymidylic acid methyl synthesis

O'Brien (15) suggests that vitamin B₁₂ is involved in the following reaction.



d. Purine biosynthesis

Luhby and Cooperman (4) postulate that vitamin B₁₂ interacts with folic acid in the formylation of AIC ribotide (AICAR) to formamidoimadazolecarboxamide ribotide (FAICAR).



AIC ribotide is an intermediate in the biosynthesis of inosinic acid, the first purine to be synthesized and from which adenine and guanine nucleotides are derived.

D. BIOCHEMICAL ASPECTS OF VITAMIN B₁₂ DEFICIENCY

A characteristic feature of vitamin B₁₂ or folic acid deficiency is the megaloblastic erythropoiesis which results in the production of abnormal erythrocytes. The megaloblast is a morphologic entity characterized by a large cell size, increased ratio of cytoplasmic to nuclear area, a characteristic fine grained chromatin texture and cytoplasmic basophilia. The megaloblast contains a normal or slightly increased amount of DNA and a substantially increased amount of RNA per cell i.e. an increased RNA/DNA ratio. This cell is in a state of unbalanced growth due to impaired deoxyribotide synthesis. DNA replication and cell division are blocked whereas synthesis of cytoplasm (RNA and protein) proceeds normally, hence the RNA/DNA ratio rises. Buchanan (16) suggests that the reactions of the methionine cycle (homocysteine - methionine reaction) are critical in the disordered erythropoiesis of folic acid and vitamin B₁₂ deficiencies. In vitamin B₁₂ deficiency, the inability to convert N⁵-methyl THF to THF results in a trapping of methyl THF (the storage form of folate) with a resultant "functional" deficiency of folate. The folate coenzymes function in the metabolism of "one-carbon" units, principally for the transfer of methyl, methylene, and formyl groups for the synthesis of compounds involved in the biosynthesis of purines and pyrimidines and hence RNA and DNA. In purine biosynthesis the reactions in which folate coenzymes are involved are the formylation

of glycinamide ribotide to formylglycinamide ribotide and the formylation of AIC ribotide to formaminoimidazole-carboxamide ribotide. The first reaction depends on $N^{10}N^5$ -methenyl THF and the second on N^{10} -formyl THF. In pyrimidine biosynthesis, the methylation of deoxyuridylic acid to thymidylic acid (the latter the pyrimidine nucleotide which differentiates DNA from RNA) requires $N^{10}N^5$ -methylene THF.

Luhby and Cooperman (4) on finding an increased urinary excretion of AIC in vitamin B_{12} deficiency suggest a block in the reaction



in the purine biosynthetic pathway which could account for the defective nucleic acid synthesis in pernicious anemia. However, it is only by inference that this may be a vitamin B_{12} dependent reaction.

The methylmalonyl - CoA isomerase reaction is not concerned with the abnormal nucleoprotein metabolism of megaloblastic anemia and it is not known what physiological consequences may result from impairment of this reaction.

Neurologic symptoms are a feature of 80 to 90% of all cases of pernicious anemia (17). No known specific biological action of vitamin B_{12} on the central nervous system has yet been shown.

The laboratory diagnosis of vitamin B_{12} deficiency includes the following procedures:

1. Hematological examination of peripheral blood and bone marrow.

2. Determination of vitamin B₁₂ concentration in the blood serum.

3. Determination of the presence of free hydrochloric acid in gastric contents.

4. Measurement of the absorption of radioactive vitamin B₁₂ (Schilling test).

There are some problems encountered in some of the above procedures. The serum vitamin B₁₂ determination usually involves a microbiological assay. The Schilling test necessitates the use of radio-active material and on occasion may give inconclusive results. It was felt, therefore, that there would be value in the development of a procedure to measure urinary MMA excretion as another index of vitamin B₁₂ deficiency.

It was proposed then to investigate the following:

1. The development of a quantitative method to demonstrate the presence of increased quantities of MMA in the urine.

2. The assessment of the excretion of MMA as a diagnostic tool for the diagnosis of vitamin B₁₂ deficiency, this to be done by comparison with the Schilling test and the hematological findings.

3. The relation, if any, of the amount of MMA excreted to the neurological symptoms present.

4. The excretion of AIC in vitamin B₁₂ deficiency.

III. EXPERIMENTAL

A. DETERMINATION OF URINARY METHYLMALONIC ACID

With the work of Barness and co-workers (3) as a basis, MMA was first determined by a paper chromatographic procedure which gave satisfactory qualitative results. As reported by Barness (3), increased amounts of MMA were observed in the urine from patients with pernicious anemia.

It was then thought that the advantages offered by thin-layer chromatography, i.e. better resolution of components, greater sensitivity, decreased time required for development, should be applied to the separation of MMA. As a result systems were developed for qualitative and semi-quantitative and finally for quantitative estimations of urinary MMA by thin-layer chromatography.

1. Paper Chromatography

The paper chromatographic procedure followed was a modification of that of Barness (3). Initially, a 5 ml. aliquot of a twenty-four hour urine collection was acidified to pH 1 with 5N H_2SO_4 and continuously extracted in a 250 ml. ether extractor with 110 ml. of ethyl ether for three hours. Subsequently, an aliquot equal to approximately 1% of the twenty-four hour urine volume (to an even 5 ml. greater than the 1% volume, i.e. 5, 10, 15 or 20 ml.) was acidified and ether extracted for from four to six hours depending on the volume. The ether extract was evaporated under a stream of air to approximately

0.5 ml. and this concentrated extract applied to a ten inch square Whatman No. 1 chromatography paper. An ascending two-dimensional system was used with ethanol : ammonium hydroxide : water (80:5:15) as the first solvent and propanol : eucalyptol : formic acid (50:50:20) and water to give persistent turbidity as the second solvent. An overnight run was used for both solvents.

After evaporation of the second solvent the chromatogram was stained with the Altman reagent (18), 10% p-dimethylaminobenzaldehyde in acetic anhydride (1 volume) and acetone (4 volumes). After drying, the chromatogram was heated at 105 - 110°C. for two to three minutes. The yellow spot of methylmalonic acid was easily identified and had an Rf of 0.30 and 0.79 in the solvent systems used.

The solvents used are those suggested by Smith (18) for organic acids and gave good separation. Barness (3) used bromcresol green as the location reagent however it was found in this laboratory that with this reagent the MMA spot was neither well defined nor easily distinguishable from the other organic acids present.

For each sample, a parallel determination was made on an aliquot of the urine to which MMA had been added.

The minimum quantity of MMA detected by the Altman reagent was determined to be approximately 100 ug. under the conditions described. Therefore, by using a minimum of 1% of the volume of urine excreted in twenty-four hours

for extraction, an excretion in excess of approximately 10 mg. or slightly less of MMA per twenty-four hours could be detected. Results were reported as "none detected" if no MMA was seen on the chromatogram or "present" if the yellow MMA spot was observed.

2. Thin-Layer Chromatography

a. Qualitative determination

Thin-layer chromatography of MMA on silica gel layers was attempted. An aqueous slurry of silica gel containing 13% CaSO_4 as a binder (Merck Silica Gel G) was spread on 20 x 20 cm. glass plates with an adjustable spreader* to give a layer 250 microns thick. Silica gel containing a fluorescent additive as well as the CaSO_4 binder (Merck Silica Gel GF₂₅₄) was also used. The plates were activated just before use by heating at 110 - 120°C, for 30 to 60 minutes.

The following solvents were tried:

1. Ethanol : Ammonia : Water (160:10:30) (18)
2. Acetonitrile : Ammonia : Water (100:5:30) (19)
3. Isopropyl ether : Formic Acid : Water (90:7:30) (20)
4. Propanol : Eucalyptol : Formic Acid (50:50:20)
plus water to persistent turbidity (18)

The following location reagents were also tried:

* Desaga Apparatus - Brinkman Instruments Incorporated.

1. Bromcresol Green (18)
2. Altman Reagent (18)
3. Potassium Ferrocyanide - Ferric Ammonium Sulfate Reagent (18)
4. Bromcresol Purple (20)

Of the solvents and location reagents used it was found that ethanol : ammonia : water as solvent in a one-dimensional system and bromcresol green as location reagent gave the most satisfactory results. In this system, MMA gave a distinct yellow spot on a blue background, well separated from other organic acids present in urine.

The initial runs were made using the Desaga tank lined with solvent saturated filter paper. The plates were equilibrated in the tank before allowing the solvent run to begin. Rather variable Rf values for MMA (0.20 to 0.30) and a variation in the time required to give a solvent front of a specific distance were observed. Subsequently the CAMAG "Sandwich" Chamber was used which gives a consistent saturated atmosphere and with which no pre-equilibration is required. With this apparatus it was found that a four hour run gave a solvent front of approximately 15 cm. and that the Rf of MMA averaged 0.42. 10 μ g. of MMA was the minimum amount that could easily be detected by this procedure using bromcresol green as location reagent.

Urine samples were ether extracted as for paper chromatography. It was found that 2 mg. of MMA was

completely extracted from 10 ml. of solution (corresponding to 200 mg. per liter) in four hours. 2 mg. of MMA was about 85% removed from 20 ml. of solution after four hours extraction but was completely removed after six hours. The length of the extraction time depended, therefore, on the volume of urine being extracted. The extract was evaporated to 0.5 ml. and amounts of this extract varying from 10 to 50 μ l. were applied to the thin-layer plate.

By application of varying amounts of a standard MMA solution and of varying quantities of the extract of an unknown urine and visual comparison of the size and the intensity of the unknown spots with those of the standards, a semiquantitative estimate of the MMA present could be made. Four standard spots equivalent to 20, 40, 60 and 80 μ g. of MMA and 10, 20, 25 and 50 μ l. of the unknown extract were routinely applied

Figure III represents a thin-layer chromatogram on which four standard spots and four amounts of a urine extract were developed.

b. Quantitative determination

First attempts at quantitation were made on extracts of the MMA-containing gel scraped off the plate. The spots were located by one of two methods. A standard MMA solution was run at the position nearest one side edge



FIGURE III. Thin-layer chromatogram of standard MMA and urine extract.

of the plate. After development, the entire plate but that area above the point where this standard had been applied was covered by a second glass plate (uncoated) and the remaining strip at the edge sprayed with bromcresol green and the MMA spot located. Areas of uniform size corresponding in position to the MMA spot were removed from the plate. Alternatively, when using the silica gel containing the fluorescent material, MMA spots could be located under short-wave ultra-violet light as spots absorbing the ultra-violet light.

To the MMA-containing gel was added water to elute the MMA. When desired the silica gel was removed by centrifugation.

Various approaches to quantitate the MMA by chemical means were attempted. These included:

1. Titration of the acid
2. Determination of acidity by iodometric titration (an acid catalyzed reaction) (21)
3. Determination of the organic acid with a copper-benzidine reagent (22)
4. Conversion of the acid to a hydroxamate and formation of the colored iron chelate (23)

All showed some promise when applied to aqueous standard solutions of MMA but none was satisfactory when applied to the MMA-containing gel or to the eluate. It was also found that the sensitivity which could be obtained with aqueous standards was not as great as would be necessary for

the amounts present in urine extracts, i.e. a minimum sensitivity of 10 - 20 μ g. MMA.

Quantitation of MMA was then attempted by means of densitometry. The problem here was the removal of the gel in an intact and more or less rigid form in order to scan it. After being sprayed with bromcresol green, the gel was sprayed with a plasticizing emulsion (Neatan, Merck) and allowed to dry. Then broad (eight inch) cellophane tape was applied directly to the gel and with care the gel could be removed from the glass plate, and the MMA containing strip scanned. The instrument used was the Chromoscan Reflectance Densitometer. Filters 5021 and 5022 (585 and 595 μ .) and a 0.5 by 20 mm. slit were used. The tracing obtained had a high baseline with depressed peaks representing the yellow spots of MMA. Plotting the area under the curve against the amount of MMA present gave an approximately straight line in the 20 to 80 μ g. region.

Application of the plasticizing emulsion had the disadvantage of causing a certain lack of uniformity of the background color and had an overall lightening effect on the gel.

At this time a commercially prepared 100 micron thick layer of silica gel containing an inert plastic binder applied to a flexible plastic support became available (Eastman Chromagram). The gel adheres quite firmly to the support and the sheet can be cut as desired

without damage to the gel layer. The Chromagram seemed most adaptable for quantitation of MMA by densitometric scanning as it eliminated the need for plastic coating the gel.

Standards and samples were run on the 20 x 20 cm. Chromogram in a manner identical to that used with the glass plates. The time for development was considerably longer, being about eight hours to give a 15 cm. solvent front as compared with about four hours for the 250 micron thick silica gel on glass plates. The Rf of MMA on the Chromagrams was about 0.45.

It was found that heating the Chromagram at 110 - 120°C. for two to four minutes after spraying with bromcresol green resulted in a clearer definition of the yellow MMA spots. A strip containing these spots was cut out and scanned. Results using the Chromagram were rather more reproducible than those using the plastic-sprayed plates. Figure IV shows a graph typical of those obtained by plotting the area under the curve against the amount of MMA present.

3. Discussion

While the Chromograms were much more suitable for the densitometric scanning, the sensitivity was somewhat less. 20 μ g. of MMA could be detected on the Chromagram with about the same ease as could 10 μ g. of MMA on the 250 micron thick layers of silica gel on glass plates. It was also found latterly that there was some variation from lot

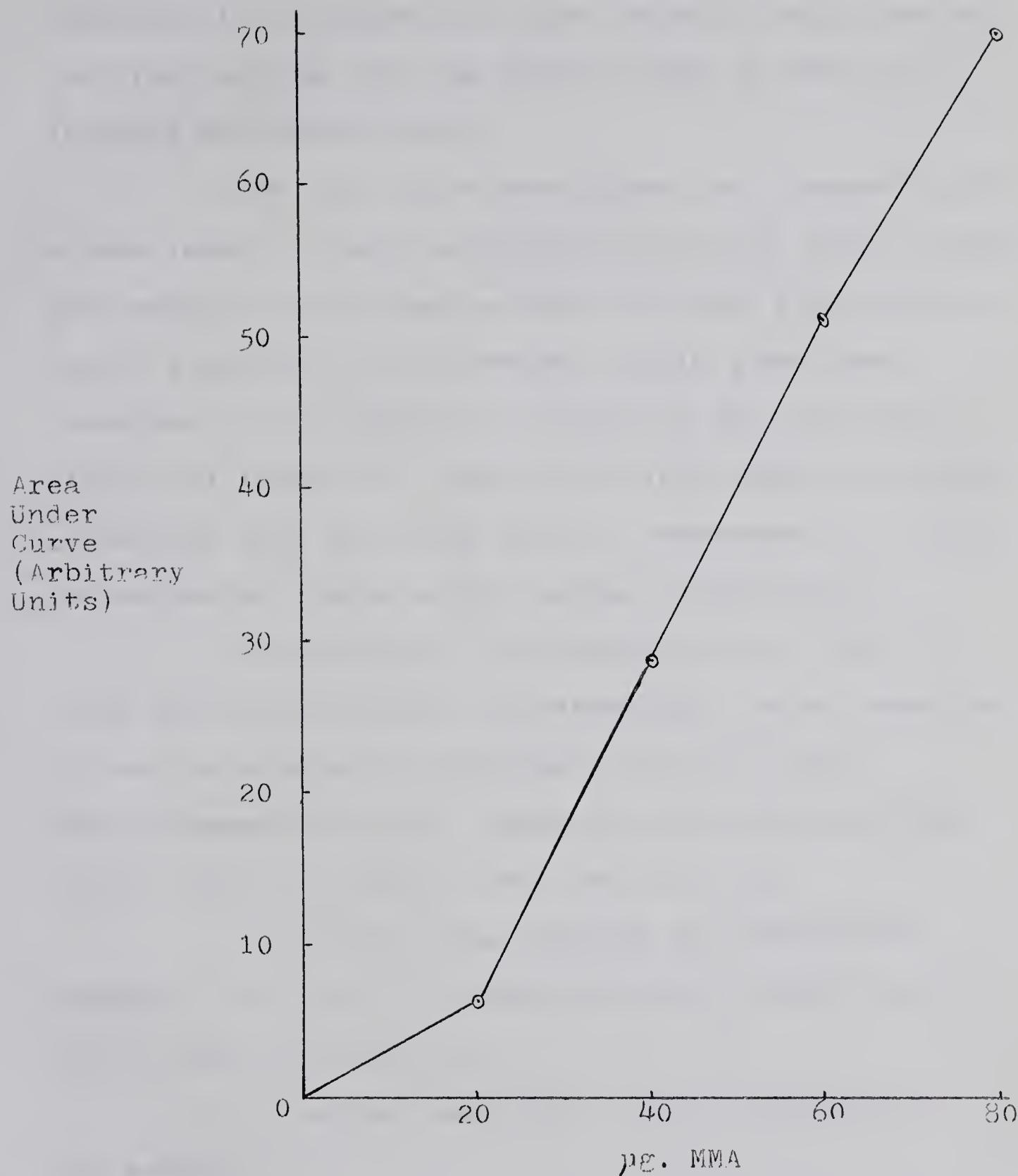


FIGURE IV. Standard curve typical of those obtained by plotting areas under the curves versus micrograms of MMA present.

to lot in the time of development required for the Chromograms. With most chromatograms, the solvent front travelled 14 to 15 cms. in eight hours but with one lot the time required for the solvent front to move this distance was twelve hours.

With both 250 micron layers and Chromogram 100 micron layers, visual comparison of unknown with standard gave results very close to those obtained from densitometric scanning, on an average, within 5 per cent. Therefore, it is probable that use of the 250 micron silica gel layer on a glass plate plus visual comparison of unknown with standards gives an assessment of urinary MMA excretion adequate for routine investigation.

In conclusion, the determination of MMA in urine by the thin-layer chromatographic method developed offered considerable improvement over the paper chromatographic method. Among the advantages offered by the thin-layer method are the following:

1. Shorter time required for development because of the use of a one-dimensional system rather than a two-dimensional one.
2. Better resolution of the components of the extract.
3. Greater sensitivity. The maximum sensitivity on the 250 micron silica gel plate using bromcresol green is 10 μ g. or less whereas on paper using the Altman reagent

it is 100 μ g, or slightly less.

4. Quantitative measurement of MMA levels in urine could be made.

4. Procedure for the determination of urinary methylmalonic acid

The following is an outline of the method used for the determination of urinary MMA in the series of patients studied.

1. Specimen: A twenty-four hour urine collection was made and an aliquot of the specimen kept frozen until used.

2. Extraction: Approximately 1% of the twenty-four hour urine volume was ether extracted as described and the extract reduced in volume to 0.5 ml.

3. Semiquantitative determination: Using a 250 micron layer of silica gel on a glass plate varying quantities of the extract were applied (up to 50 μ l.) and a visual comparison with standards after spraying with bromcresol green gave a semiquantitative estimate of the quantity of MMA present. When 50 μ l. of a comparable extract of a 1% volume of normal urine is applied, no MMA can be detected.

4. Quantitative determination: Here the Chromagram was used as previously described and the MMA containing strip scanned on the Chromoscan. The volumes of unknown extract to be applied were determined from the results of the 250 micron thick silica gel plate and were

chosen such that they contained between 20 and 80 μ g. of MMA.

Twenty-one samples were run in duplicate including duplicate extractions. The standard deviation based on the duplicate samples run was \pm 9 mg. per twenty-four hours as calculated by the method of Youden (24), with a mean of 218 mg. per twenty-four hours. This gave a coefficient of variability of \pm 4%.

B. DETERMINATION OF URINARY AIC

The method used for the measurement of urinary AIC was a slight modification of that of McGeer et al (25) and is as follows.

125 ml. of a twenty-four hour urine collection is saturated with salt, acidified to about pH 2 and the volume measured to record the increase due to salt saturation. This urine is extracted with five 40 ml. volumes of ethyl acetate. Two 50 ml. aliquots of the aqueous phase are made alkaline to pH paper with a 1 : 4 mixture of concentrated ammonium hydroxide and pyridine. To one is added 50 μ l. (25 μ g.) of a 60 mg./100 ml. solution of AIC to give a parallel analysis for recovery of AIC.

Both aliquots are extracted for three minutes with nine 26 ml. volumes of a 100 : 5 n-butanol - pyridine mixture and the organic extracts for each combined. The organic phases are acidified by the addition of 10 μ l. of concentrated sulfuric acid and shaken for three minutes.

The aqueous layers containing the AIC are removed and the butanol layers washed once with 10 ml. of 0.2N H_2SO_4 then six times with 5 ml. of 0.2N H_2SO_4 . The washings are added to the aqueous phase to give a final volume of about 40 ml.

Two 2 ml. aliquots of each extract are analyzed colorimetrically for non-acetylable diazotizable amines by coupling with N(1-naphthyl)-ethylene diamine. To the first aliquot in a tube of 22 mm. diameter is added 0.1 ml. of acetic anhydride and to the second 0.3 ml. of acetic anhydride. After vigorous shaking the tubes are allowed to stand for twenty minutes. 3 ml. of 0.2N H_2SO_4 is added to the first tube and 2.8 ml. to the second and then 0.5 ml. of 0.1% sodium nitrite is added to both. Exactly two minutes later, 0.5 ml. of 0.5% ammonium sulfamate is added followed immediately by 0.5 ml. of 0.1% N(1-naphthyl)-ethylene diamine. The mauve color which develops is read at 540 μ . at ten minute intervals until a peak reading is obtained (usually within one hour). An extract blank is also run which is exactly comparable except that the N(1-naphthyl)-ethylene diamine is omitted and 0.5 ml. distilled water added instead.

Standards containing 3, 6, 9 and 12 μ g. AIC (0.5, 1.0, 1.5 and 2.0 ml. of a 0.6 mg./100 ml. solution of AIC in 0.2N H_2SO_4 plus 0.2N H_2SO_4 to give a total volume of 2 ml.) are similarly analyzed and read at ten minutes against a reagent blank.

Quantities of AIC in the extracts are obtained by comparison with the standards. Appropriate calculations give the twenty-four hour urinary excretion of AIC. Normal values as quoted by McGeer are 0.98 ± 1.16 (2 S.D.) mg./day with a range of 0.3 to 2.5 mg./day.

IV. CLINICAL STUDIES

A. METHYLMALONIC ACID

1. Introduction

Urinary MMA excretion was determined on a total of 91 patients with a variety of clinical conditions. Many had findings which pointed to a possible deficiency of vitamin B₁₂. The results obtained are tabulated in Table I with detailed information on patients with laboratory or clinical evidences indicative of vitamin B₁₂ deficiency. In addition other pertinent laboratory findings, where available, are included.

In the early patient studies, the MMA excretion was determined only by paper chromatography. These results are recorded as "positive" (MMA present) or "negative" (no MMA detected). Subsequently, thin-layer chromatography was used in addition to, or instead of, paper chromatography. These results are recorded as "less than" a certain number of milligrams per twenty-four hours rather than negative or as a quantitative result. It was found that so called "negative" results (the normal situation) represented values of less than 10 mg. MMA per twenty-four hours.

Serum vitamin B₁₂ levels were determined by microbiological assay with the use of *Euglena gracilis*, as described by Ross (26). Normal levels are from 140 to 520 μ g. per ml. Serum folic acid levels were measured by the method of Herbert (27) using *Lactobacillus casei*. Normal serum folic acid levels are in excess of 3 μ g. per ml. Qualitative urinary formiminoglutamic acid (FIGLU) after a loading dose of 15 g. L-Histidine was determined by the method of Kohn et al (28). The measurement of urinary FIGLU after histidine loading recently has been studied as an indicator of folic acid deficiency. The conversion of FIGLU, a metabolite of histidine, to glutamic acid involves the transfer of the formimino group to tetrahydrofolic acid. Thus, in a deficient state, the transfer reaction is diminished with the subsequent excretion of increased amounts of FIGLU ("positive" result). In the normal condition, no FIGLU is detected ("negative" result). Co⁶⁰-labelled vitamin B₁₂ absorption tests (Schilling tests) were performed using 0.5 μ c. Co⁶⁰-vitamin B₁₂. Normal urinary excretion is in excess of 6% of the oral dose in twenty-four hours. The other laboratory tests, hemoglobin, hematological examination (peripheral blood and bone marrow smears) and gastric analyses were performed by standard procedures.

From one patient who was excreting increased quantities of MMA, three consecutive eight hour collections were taken in order to observe whether or not the MMA

TABLE I

Laboratory data of patients studied

Patient	Sex	Age	Hemo-globin g./ 100 ml.	Hematological Findings	Free Gastric HCl	Urine FIGLU	Serum Folic Acid mpg./ ml.	Serum Vit. B ₁₂ μg./ ml.	Co ⁶⁰ B ₁₂ Absorption with out out IF %	Urine IF %	Urine MMA mg./ 24 hr.	Urine AIC mg./ 24 hr.	Neuro- logical symptoms	Diagnosis
# 1	F	48	6.8	macrocytic megaloblastic anemia	neg	---	---	---	1.2	---	146	0.9	slight	pernicious anemia
# 2	F	52	12.9	normal	neg	---	---	---	0.9	1.0	pos	---	severe	pernicious anemia
# 3	M	84	13.7	normal	neg	---	---	---	4.5	---	4.1	neg	---	severe
# 4	F	68	7.1	megaloblastic anemia	neg	---	---	---	4.0	0.8	6.1	36	0.7	normal
# 5	F	56	5.9	macrocytic megaloblastic anemia	neg	---	---	---	3.5	0.2	3.1	34	1.1	---
# 6	M	74	8.0	macrocytic megaloblastic anemia	neg	pos	---	---	5.5	0.2	---	73	1.1	slight
# 7	M	84	2.3	macrocytic megaloblastic anemia	---	---	---	---	6.0	---	---	pos	---	slight

TABLE I (cont'd)

Patient	Sex	Age	Hemoglobin g./ 100 ml.	Hematological Findings	Free Gastric HCl	Urine FIGLU	Urine Vit. B12 μg./ ml.	Serum Folic Acid μg./ ml.	Co ⁶⁰ B ₁₂ Absorption with out IF %	Urine MMA mg./ 24 hr.	Urine AIC mg./ 24 hr.	Neuro- logical symptoms	Diagnosis		
#	S	190	8.2	moderate macrocytic normoblastic anemia	neg	pos	---	---	0.3	---	53	1.1	normal	pernicious anemia	
#	9	F51	14.3	normal	neg	---	---	---	0.8	5.7	1680	0.8	severe	pernicious anemia	
#	10	M73	11.2	normal	neg	---	---	---	0.7	5.8	620	1.8	slight	pernicious anemia	
#	11	F86	12.7	macrocytic anemia	---	---	---	---	40	2.5	14	77	0.5	normal	pernicious anemia
#	12	M75	---	macrocytic anemia	---	---	---	---	0.25	---	60	0.45	---	pernicious anemia	
#	13	M71	6.4	megaloblastic erythropoiesis	---	---	5.7	43	0.1	---	106	0.4	---	pernicious anemia	
#	14	M68	9.5	macrocytic - slight megaloblastic erythropoiesis	---	7.0	---	---	1.5	---	150	1.4	normal	pernicious anemia	

TABLE I (cont'd)

Patient	Hemo- globin g./ 100 ml.	Hematological Findings	Free Gastric HCl	Serum Vit. B ₁₂ μg./ ml.	Urine VITLU	60 B ₁₂ Absorption with out IF %	Urine MMA mg./ 24 hr.	Urine AIC mg./ 24 hr.	Neuro- logical symptoms	Diagnosis	
# 15	M 3	5.9	macrocytic megaloblastic anemia	pos	24	4.0	3.4	4.5	280	0.15	normal pernicious anemia of infancy
# 16	M 4	9.1	macrocytic megaloblastic anemia	---	---	1.9	4.0	1.5	---	2.5	normal pernicious anemia
# 17	M 2	6.4	macrocytic megaloblastic anemia	---	---	7.6	<20	3.2	---	736	3.0 slight pernicious anemia
# 18	M 4	7.9	moderate macrocytic megaloblastic anemia	neg	5.3	<25	0.2	---	75	0.9 slight pernicious anemia	
# 19	M 2	14.1	slight macrocytosis & hypersegmenta- tion	neg	4.7	65	0.5	5.0	45	1.8 severe pernicious anemia	
# 20	F 51	7.3	moderate pancytopenia	neg	>40	<20	0.7	---	47 (5 hr.)	10.0 severe pernicious anemia	
# 21	M -	---	megaloblastic anemia	---	5.5	<60	0.7	---	12	1.2 ---	

TABLE I (cont'd)

Patient	Sex	Age	Hemo- globin g./ 100 ml.	Hematological Findings	Free Gastric HCl	Urine FIGLU	Serum Vit. B ₁₂ μg./ ml.	Co ⁶⁰ B ₁₂ Absorption with out IF %	Urine MMA mg./ 24 hr.	Urine AIC mg./ 24 hr.	Neuro- logical symptoms	Diagnosis	
# 22	F	60	5.5	macrocytic megaloblastic anemia	neg	pos	8.0	<20	2.6	6.0	12	1.4	normal pernicious anemia
# 23	F	80	---	macrocytic megaloblastic anemia	---	---	20	2.9	---	186	0.85	---	pernicious anemia
# 24	M	77	11.6	macrocytic partially megaloblastic anemia	neg	---	---	<40	2.6	---	55	1.6	slight pernicious anemia
# 25	F	68	14.4	normal	neg	---	---	---	0.35	1.5	neg	---	slight diabetes mellitus
# 26	M	52	15.6	normal	pos	---	---	---	---	2.5	1.7	neg	---
													cervical spondylosis ?? vit. B ₁₂ deficiency

TABLE I (cont'd)

Patient	Sex Age	Hemo- globin g./ 100 ml.	Hematological Findings	Free Gastric HCl	Serum Vit. B12 μ g./ ml.	Co ⁶⁰ B ₁₂ Absorption with IF out IF % %	Urine MMA mg./ 24 hr.	Urine AIC mg./ 24 hr.	Neuro- logical symptoms	Diagnosis
# 27	F'73	11.5	normal	---	---	2.4	---	< 8	2.7	---
# 28	M'70	8.5	moderate hypochromic anemia	neg	6.1	65	5.3	---	1.1	normal
# 29	M'81	12.7	normal	---	2.6	45	3.4 (vol. low)	---	---	carcinoma of stomach - inoperable
# 30	M'36	11.4	non-chromic zone cortosis	---	1.4	275	6.2	---	< 12	6.5
					nos					celiac sprue secondary osteomalacia iron & folic acid deficiency anemia

TABLE I (cont'd)

Patient	Sex	Age	Hemo- globin g./ 100 ml.	Hematological Findings	Free Gastric HCl	Urine FIGLU	Urine Vit. B ₁₂ μg./ ml.	Serum Folic Acid μg./ ml.	Co ⁶⁰ B ₁₂ Absorption with out out IF IF IF %	Urine IF IF IF %	Urine AIC mg./ 24 hr.	Urine TMA mg./ 24 hr.	Neuro- logical symptoms	Diagnosis	"miscellaneous" patients
# 31	M	19	11.8	slight to moderate hypochromia	---	pos	6.6	418	3.1	---	<10	5.0	normal	idiopathic sprue diabetes mellitus	
# 32	M	16	11.6	slight normoblastic hyperplasia	---	---	---	---	11.5	---	<10	5.2	normal	aplastic anemia diffuse interstitial fibrosis	<10 normal in 29 patients

TABLE II

Urinary MMA levels in three consecutive
eight hour collections from one patient

<u>Patient</u>	<u>Time</u>	<u>MMA mg./24 hr.</u>
#24	12 midnight - 8 am.	26
	8 am. - 4 pm.	13
	4 pm. - 12 midnight	17

TABLE III

Urinary MMA levels before and after
vitamin B₁₂ therapy

<u>Patient</u>	<u>Day</u>	<u>Intramuscular Dose of Vit. B₁₂</u>	<u>MMA mg./24 hrs.</u>
# 9	1	--	1680
	4 months	after repeated doses of 1000 μ g.	< 10
#16	1	--	260
	2	1000 μ g.	
	3	400 μ g.	
	5	400 μ g.	
	7	400 μ g.	
	12	400 μ g.	
	13	--	< 10
#17	1	--	736
	3	1000 μ g.	
	8	500 μ g.	
	10	--	10
#24	1	--	54
	2	--	56
	5	1000 μ g.	
	9	--	19
	12	1000 μ g.	
	13	--	21
	14	1000 μ g.	
	16	1000 μ g.	
	19	--	< 5

excretion varied throughout the day. These results are shown in Table II.

On four patients MMA excretion was again measured after injections of vitamin B₁₂ had been given. Specimens were collected from one of these patients at frequent intervals until MMA had disappeared from the urine. Table III gives these results.

2. Results

In Table I are the results obtained from a study of twenty-four patients on whom a diagnosis of pernicious anemia was made. Of this group, sixteen out of twenty-one patients tested had lower than normal hemoglobin levels and nineteen of the twenty-four showed macrocytosis or megaloblastic erythropoiesis on the peripheral blood or bone marrow film. Serum vitamin B₁₂ levels determined on eighteen of the patients were well below the lower limits of normal. Co⁶⁰-vitamin B₁₂ absorption as done on twenty-two of the patients was also much reduced. Histamine-fast achlorhydria was a finding in sixteen of seventeen patients tested, the one patient showing free gastric acid being a three year old with a diagnosis of "pernicious anemia of infancy".

Urinary methylmalonic acid excretion on all but one of the patients was found to be elevated. This patient, #3, had been under treatment for pernicious anemia until two years prior to being investigated early in this study. His serum vitamin B₁₂ level was low, 45 μ g. per ml., his vitamin B₁₂ absorption even with intrinsic factor was

slightly low and his urinary MMA excretion was "negative". In twenty-one of the remaining twenty-three patients of this group on whom urinary MMA was quantitated, the twenty-four hour excretion ranged from 10 to 1,680 mg., the normal excretion being less than 10 mg. per day.

Patients #25 to #29 had one or more laboratory findings which suggested vitamin B₁₂ deficiency. Patients #25 and #26 had a decreased vitamin B₁₂ absorption which did not increase when repeated with intrinsic factor. Patient #25 had a secondary diagnosis of vitamin B₁₂ deficiency while for patient #26 the same diagnosis was queried. The urinary MMA excretion in both these patients was "negative" as determined early in the series by qualitative paper chromatography prior to the development of the thin-layer quantitative method. Both patients had a normal hematological picture.

Patients #27 and #28 had a diagnosis of mal-absorption. Patient #27 had a low vitamin B₁₂ absorption. Patient #28 had a normal vitamin B₁₂ absorption but a low serum vitamin B₁₂ level. In both cases MMA excretion was normal. Patient #29, diagnosed as inoperable carcinoma of the stomach, had a low serum vitamin B₁₂ level but a "negative" MMA excretion.

Patients #30, #31 and #32 are included in Table I as they had elevated urinary AIC levels. These patients will be discussed later.

The remaining fifty-nine cases represent hospital patients from medical and surgical wards with a wide variety of disorders other than vitamin B₁₂ deficiency and make up the "miscellaneous" group in Table I. Included in this group were patients with iron and folic acid deficiency anemias, malabsorption showing no evidence of vitamin B₁₂ deficiency, pneumonia and many other conditions. This group was assessed to study the specificity of urinary MMA excretion as an index of vitamin B₁₂ deficiency and to observe any false positive results in disorders not related to a deficiency of vitamin B₁₂. No increase in MMA excretion above normal was observed in this group.

The neurological symptoms have been tabulated in Table I in three rather broad groups; normal, slight (including minor neurological involvement such as numbness and/or tingling in the extremities) and severe (being subacute combined degeneration of the spinal cord).

Table II shows the eight-hourly MMA excretion of the one patient studied to observe any variation within the day in urinary MMA. In this particular patient the overnight collection showed a greater amount of MMA excreted than either of the other two eight-hour periods neither of which differed appreciably from the other.

As is shown in Table III, MMA excretion returns to normal levels within a few days of the patient receiving intramuscular injections of vitamin B₁₂. Patient #9 was not followed closely but after a four month period during

which she received a series of intramuscular injections of vitamin B₁₂, the twenty-four hour MMA excretion dropped from 1,680 mg. to less than 10 mg. In the other three cases, the MMA excretion returned to normal within ten days to two weeks.

3. Discussion

All but one of the patients on whom a diagnosis of pernicious anemia was established showed an elevation of urinary MMA. As has been stated, the one patient on whom MMA was recorded as "negative" was assessed using the paper chromatographic procedure and may possibly have shown a slightly increased level of MMA which would not have been detected under the early and less sensitive conditions of this procedure. In all other cases of pernicious anemia, MMA excretion served equally as well as the Co⁶⁰-vitamin B₁₂ absorption as a measure of vitamin B₁₂ deficiency. Like labelled B₁₂ absorption, urinary MMA may be abnormal before there is hematological evidence of the vitamin deficiency. This is evidenced by those patients in the series who showed no anemia or megaloblastic erythropoiesis yet had decreased vitamin B₁₂ absorption and increased MMA excretion.

In considering the urinary MMA levels and the neurological symptoms shown by the patients with pernicious anemia, there would appear to be no correlation between the two. There is no method for determining blood MMA, but assuming MMA excretion to reflect body levels of MMA, an

increased quantity of circulating MMA apparently does not relate to the degree of neurological involvement. One patient with subacute combined degeneration of the spinal cord had an MMA level of 1,680 mg. per twenty-four hours while another with the same condition had an MMA level of 45 mg. per twenty-four hours. On the other hand, patients with no neurological disorders had MMA levels ranging from 12 to 280 mg. per twenty-four hours. This is consistent with previous findings that other measures of vitamin B₁₂ deficiency do not correlate with the degree of neurological involvement.

A sodium propionate loading test was considered on the basis that biochemically speaking, this could be a useful procedure for revealing mild deficiencies of body levels of vitamin B₁₂ that might not appear from clinical and other observations. However, the reports of White and Cox (2) and of Kahn and coworkers (29) in the very recent literature (July, 1965) of attempts using this approach were not encouraging so this was not carried out.

While no conclusions can be drawn from the results on only one patient, the variation in MMA excretion in consecutive eight hour specimens in this patient is in agreement with the observations of Kahn et al (29). This group found there could be considerable variation in the amount of MMA excreted from one eight hour period to the next.

The observed drop in MMA excretion after administration of vitamin B₁₂ to the patient is an expected finding and demonstrates that it is biologically valid to relate MMA excretion and B₁₂ levels and that increases are not merely spurious findings.

B. 4(5)-AMINO-5(4)-IMIDAZOLECARBOXAMIDE

1. Introduction

Urinary AIC was determined on forty-nine patients, twenty of whom had a diagnosis of pernicious anemia. The results on those patients with pernicious anemia are included in Table I along with those on four other patients who showed an increased level of urinary AIC. AIC was also determined on three of the patients on whom MMA estimations were done after treatment with vitamin B₁₂. These results are in Table IV.

2. Results

Of the group of twenty patients with pernicious anemia, only three had elevated levels of urinary AIC. Among the remaining twenty-nine cases, four had higher than normal levels of AIC. Three of these four patients suffered from malabsorption while the fourth had an aplastic anemia.

The three patients recorded in Table IV all showed an appreciable drop in their urinary AIC levels after vitamin B₁₂ therapy.

TABLE IV

AIC excretion before and after treatment with
intramuscular injections of vitamin B₁₂

<u>Patient</u>		<u>AIC mg./24 hrs.</u> (Normal <2.1 mg./24 hrs.)
#16	Before	2.5
	After	1.0
#17	Before	3.0
	After	0.8
#24	Before	1.6
	After	0.9

3. Discussion

The results of this study on twenty patients do not show increased urinary AIC excretion to be a constant feature of pernicious anemia and reduced B_{12} levels as evidenced from depressed isomerization of methylmalonyl-CoA to succinyl-CoA. If indeed vitamin B_{12} is directly involved in the folic acid dependent formylation of AICAR to FAICAR rather than merely being necessary for the primary activity of folic acid, this enzymatic conversion is still operative at low levels of B_{12} availability and in the presence of clinical evidence of B_{12} deficiency. Thus AIC excretion does not appear to be a reliable measure of B_{12} deficiency. However, it should be noted that other authors studying smaller series of patients with B_{12} deficiency have shown an increased AIC excretion. Luhby and Cooperman (4) on finding urinary AIC to be elevated in vitamin B_{12} deficiency states but not in folic acid deficiencies interpreted this as indicating that deficiency of vitamin B_{12} alone specifically results in increased AIC excretion. Herbert (5) and Middleton (6) found increased urinary AIC in both B_{12} and folate deficiencies and these authors suggest that folic acid deficiency alone produces the abnormally high urinary excretion of AIC. They conclude that as vitamin B_{12} is thought to be necessary for the normal activity of folic acid, the similar AIC findings in vitamin B_{12} deficiency alone are not unexpected. It is interesting that in those

patients on whom AIC was measured before and after they received vitamin B₁₂ there is an appreciable drop in AIC excretion after treatment. Thus on this basis it would appear that there is some validity to the proposal that B₁₂ is involved either primarily or secondarily in the formylation of AICAR to FAICAR.

V. CONCLUSION

1. A quantitative method for the determination of MMA in urine has been developed which uses the technique of thin-layer chromatography following an ether extraction of urine. By this method normal urinary excretion of MMA is less than 10 mg. per day and the coefficient of variability is four percent.

2. In twenty-three cases of clinically proven pernicious anemia, the urinary MMA excretion was increased in all cases. Thus, measurement of MMA excretion is a sensitive index of vitamin B₁₂ deficiency. Urinary MMA returns to normal in response to vitamin B₁₂ therapy.

3. All cases with a proven diagnosis of pernicious anemia and low Co⁶⁰-vitamin B₁₂ absorption (Schilling test) also exhibited increased urinary MMA levels. Thus for laboratories not licensed for use of radioisotopes necessary for the Schilling test, urinary MMA measurement should be considered to be a reliable and adequate substitute in the laboratory diagnosis of pernicious anemia.

4. The degree of neurological involvement in pernicious anemia appears to be independent of the elevated

body levels of MMA as evidenced by MMA excretion.

5. The excretion of MMA in patients with pernicious anemia may be elevated before anemia or the abnormal hematological features develop. This aspect should be an aid to the early diagnosis of pernicious anemia.

6. Urinary excretion of MMA in successive eight hour periods as measured in one patient was variable. This finding concurs with reports of others who have studied the diurnal excretion of MMA.

7. Urinary AIC was only occasionally increased in those patients with pernicious anemia and the study does not give any further information on the possible direct involvement of vitamin B₁₂ in the reaction AICAR to FAICAR, a reaction involved in the synthesis of the nucleotide, inosinic acid. Although it is well established that vitamin B₁₂ deficiency leads to disordered nucleic acid synthesis, results in this study do not show clearly the involvement of vitamin B₁₂ in the AICAR to FAICAR step.

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